

Oxalic acid/oxalates in plants: from self-defence to phytoremediation

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Oxalic acid and oxalates are produced and present in plants in different amounts. Insoluble calcium oxalate plays a key role in regulating calcium concentration, which is important in the functioning of guard cells. Oxalates provide tolerance to aluminium toxicity to plants growing in acid soils. Both oxalic acid and calcium oxalate provide self-defence against insect pests and grazing animals. Oxalates are involved in phytoremediation of soils rendered toxic by heavy metals, like lead, cadmium, zinc, etc.

Keywords: Aluminium toxicity, calcium oxalate, oxalic acid, phytoremediation.

Oxalic acid and oxalates are present in leaves, roots, stems, fruits and seeds of many plants. Oxalic acid is a dicarboxylic acid with the formula [HOOC.COOH] or H₂C₂O₄. Its acidic strength is greater [pKa = 1.27 for the dissociation of the first H⁺] than that of acetic acid [pKa = 4.76]; it is probably the strongest organic acid in plants. It is a reducing agent and its conjugate base [C₂O₄]²⁻ is a chelating agent for cations such as Ca, Mg, Zn, Mn, Fe, etc.¹. Oxalic acid reacts with cations resulting in the formation of different oxalates. The solubility in water (20°C/25°C) of oxalic acid is 143 g l⁻¹, while that of sodium, potassium and magnesium oxalates is 37.0, 39.3 and 1.2 g l⁻¹ respectively. In contrast, the solubility of calcium oxalates is only 0.67 mg l⁻¹. Because of its metallic ion-chelating properties, oxalic acid is widely used for bleaching purposes in dyeing industry, especially for removing rust spots. It is also used for the restoration of old wood.

Production of oxalic acid in plants

Oxalic acid was isolated from the extract of a rhizomatous plant wood sorrel (*Oxalis acetosella*) as early as 1773 by Francois Pierre Savary in Switzerland². Oxalates may constitute as much as 3–10% of plant dry mass³. Three pathways for oxalate biosynthesis in plants have been suggested, namely oxidation of glycolate/glyoxylate during photorespiration³, oxaloacetate breakdown presumably catalysed by an oxaloacetase^{4,5} and from

ascorbic acid breakdown^{3,6}. However, the relative contribution of these pathways is controversial. For example, Xu *et al.*⁷ observed that oxalate accumulation and regulation are independent of glycolate oxidation in rice, while Yu *et al.*⁸ observed that glyoxylate rather than ascorbate are an efficient precursor for oxalate biosynthesis.

Functions in physiological processes

It is suggested that oxalate synthesis is related to the regulation of the balance between inorganic cations (K⁺, Na⁺, NH₄⁺, Ca²⁺, Mg²⁺) and anions (NO₃⁻, Cl⁻, H₂PO₄⁻, SO₄²⁻) in plant cells⁹. Formation of insoluble calcium oxalate enables plants to regulate the concentration of calcium and oxalic acid, both of which can be toxic when in excess¹⁰. For example, Ca is well known to play an important role in signal transduction in stomatal guard cells, but a higher than desired concentration of Ca inhibits this function. Regulation of Ca by calcium oxalate was shown by Ruiz and Mansfield¹¹ in the case of *Cummlina communis* L. Insoluble calcium oxalate formation is suggested as a mechanism of storing Ca for future needs of the plant^{12,13}.

Protection from insect pests and grazing animals

Oxalic acid and oxalates provide biochemical as well as mechanical defence against insect pests and animals^{3,14,15}. Leaves of non-edible plants of *Arisaema* sp. (known as cobra lily) found in China and Japan, contain very high levels of calcium oxalate and are toxic. Soluble oxalates are reported to be toxic to plant hoppers¹⁶ and other insects. High oxalate-containing grasses are reported to be toxic to grazing cattle^{17,18}. For ruminants, Sidhu *et al.*¹⁹ reported a critical limit of 3.01% oxalate in Napier grass (*Pennisetum purpurea*). Death of a large number of sheep due to oxalate toxicity by grazing on poisonous weed *Halogeton glomeratus* has been reported²⁰. Calcium oxalate is fairly toxic and even a small dose is enough to cause sensation of burning in the mouth, and swelling and choking of throat. With regards to insoluble oxalate, some plants such as *Tragia ramosa*²¹ and *Medicago truncatula*²² produce needle-shaped hairs, which can puncture the dermis of insects and thus keep them away.

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Presence of calcium oxalate minerals (also referred to as biominerals) whewellite and weddellite has been reported in several species of cacti (*Opuntia* species)^{23,24}. Weddellite druses are made up of tetragonal crystallites, whereas those of whewellite have acute points and general star-like shape. Keeping this in view, Nakata²⁵ has suggested genetically engineered formation of calcium oxalate in plants to reduce insect damage.

As a chemical secreted by fungi to infect plants

Fungi such as *Sclerotium rolfsii* Sacc²⁶ and *Sclerotinia sclerotiorum*²⁷ secrete oxalic acid to infect the plants. Oxalic acid acts as a signalling molecule to induce a genetically regulated apoptotic-like programmed cell death (PCD) in host plant tissue²⁸. Thus, the fungus tricks the host into generating nutrient-rich dead cells that are of sole and direct benefit to it²⁹.

Overcoming aluminium toxicity

Acid soils account for approximately 40% of the arable land on earth. Exchangeable aluminium (Al³⁺) in soil solution has been identified as a major cause for soil acidity³⁰ and its toxicity is the limiting factor for crop productivity on such soils^{31,32}. Some crops such as cotton (*Gossypium* sp.), sorghum (*Sorghum bicolor*) and alfalfa (*Medicago sativa*) are sensitive to Al toxicity and need liming to nearly zero level of Al-saturation for successful crop growth³³. Morita *et al.*^{34,35} demonstrated that oxalate was a key compound in the Al-tolerance mechanism employed by the tea plant, which detoxifies Al³⁺ externally in the rhizosphere. Similarly, oxalic acid is secreted from the roots of buckwheat (*Fagopyrum esculentum* Moench, cv. Jianxi)^{36,37}, which also shows high Al-tolerance.

Role in phytoremediation

Oxalic acid is also reported to help in the accumulation of heavy metals, cadmium, nickel, zinc, etc. by hyper-accumulators³⁸⁻⁴¹, which are being utilized in phytoremediation of soils affected by toxicity of these heavy metals. This toxicity in soils may be caused by the continuous use of sewage sludge⁴² or closeness to zinc or lead smelters⁴³⁻⁴⁵ or due to other natural causes. About 400 species of 45 plant families, including Brassicaceae, Fabaceae, Euphorbeaceae, Asteraceae, Lamiaceae and Scrophulariaceae have been identified as hyper-accumulators of heavy metals⁴⁶. A study in Macedonia recommended alfalfa for the phytoremediation of soils made toxic with lead, oilseed rape (*Brassica napus*) and white clover (*Trifolium repens* L.) for toxicity with cadmium, and alfalfa and white clover for toxicity with

zinc⁴⁵. Of special interest is arsenic toxicity of groundwater in eastern India and Bangladesh^{47,48} and several other countries of the world⁴⁹. Large intake of arsenic results in lung, kidney and bladder cancer and several other ailments⁵⁰. Recently, Chintakovid *et al.*⁵¹ from Thailand reported that Nugget marigold, a tetraploid hybrid between American *Tagetes erecta* L. and French *T. patula* L, could be a good phytoremediator for arsenic; arsenic content was 46.2% in the leaves and 5.8% in the flowers.

Conclusion

Oxalic acid and oxalates play an important role in maintaining the desired Ca concentration in cells, self-defence against insect pests and grazing animals and thus phytoremediation, and thus deserve further attention.

1. Ullmann's *Encyclopedia of Industrial Chemistry*, Wiley-VCH, 2005, pp. 17624/28029.
2. Gmelin, L. and Watts, H., *Handbook of Chemistry*, Cavendish Society, London, UK, 1855, vol. 9, p. 111.
3. Franceschi, V. R. and Nakata, P. A., Calcium oxalate in plants: formation and function. *Annu. Rev. Plant Biol.*, 2005, **56**(1), 41–71.
4. Chang, C. C. and Beevers, H., Biogenesis of oxalate in plant tissues. *Plant Physiol.*, 1968, **43**, 1821–1828.
5. Duttan, C. M. V. and Evans, C. S., Oxalate production by fungi: its role in pathogenicity and ecology in the soil environment. *Can. J. Microbiol.*, 1996, **42**, 881–895.
6. Kostman, T. A., Tarlyn, N. M., Loewus, F. A. and Franceschi, V. R., Biosynthesis of l-ascorbic acid and conversion of carbons 1 and 2 of l-ascorbic acid to oxalic acid occurs within individual calcium oxalate crystal idioblasts. *Plant Physiol.*, 2001, **125**, 634–640.
7. Xu, H. W., Ji, X. M., He, Z. H., Shi, W. P. H., Niu, J. K., Li, B. S. and Peng, X. X., Oxalate accumulation and regulation is independent of glycolate oxidase in rice leaves. *J. Exp. Bot.*, 2006, **57**, 1899–1908.
8. Yu, L. *et al.*, Glyoxylate rather than ascorbate is an efficient precursor for oxalate biosynthesis in rice. *J. Exp. Bot.*, 2010, **61**(6), 1625–1634.
9. Calistan, M., The metabolism of oxalic acid. *Turk. J. Zool.*, 2000, **24**, 103–106.
10. Webb, M. A., Cavaletto, J. M., Carpita, N. C., Lopez, L. E. and Amott, H. J., The intravacuolar organic matrix associated with calcium oxalate crystals in the leaves of *Vitis*. *Plant J.*, 1995, **7**, 633–648.
11. Ruiz, L. P. and Mansfield, T. A., A postulated role for calcium oxalate in the regulation of calcium ion in the vicinity of stomatal guard cells. *New Phytol.*, 1994, **127**, 473–781.
12. Helper, P. K. and Wayne, R. O., Calcium and plant development. *Annu. Rev. Plant Physiol.*, 1985, **36**, 397–439.
13. Franceschi, V. R., Calcium oxalate formation is a rapid reversal process in *Lemna minor*. *Protoplasma*, 1989, **148**, 130–137.
14. Korth, K. L. *et al.*, *Medicago truncatula* mutants demonstrate the role of plant calcium oxalate crystals as an effective defense against chewing insects. *Plant Physiol.*, 2006, **141**, 188–195.
15. Nakata, P. A., Plant calcium oxalate crystal formation and its impact on human health. *Front. Biol.*, 2012, **7**(3), 254–266.
16. Yoshihara, T., Sogana, K., Pathak, M. D., Juliano, B. O. and Sakamura, S., Oxalic acid as a sucking inhibitor of the brown

- plant hopper (Delphacidae, Homoptera). *Entomol. Exp. Appl.*, 1980, **27**, 149–152.
17. Libert, B. and Franceschi, V. R., Oxalate in crop plants. *J. Agric. Food Chem.*, 1987, **35**, 926–938.
 18. Rahman, M. M., Abdullah, R. B. and Wan Khadijah, W. E., A review of oxalate poisoning in domestic animals: tolerance and performance aspects. *J. Anim. Physiol. Anim. Nutr.*, 2013, **97**(4), 605–614.
 19. Sidhu, P. K., Joshi, D. V. and Srivastava, A. K., Oxalate toxicity in ruminants fed over grown Napier grass (*Pennisetum purpurea*). *Indian J. Anim. Nutr.*, 1996, **13**, 181–183.
 20. James, P. A., Halogeton poisoning in livestock. *J. Nat. Toxins*, 2012, **8**, 395–403.
 21. Thurston, E. L., Morphology, fine structure and ontogeny of the stinging emergence of *Tragia ramosa* and *T. saxicola* (Euphorbiaceae). *Am. J. Bot.*, 1976, **63**, 710–718.
 22. Park, S. H., Doege, S. J., Nakata, P. A. and Korth, K. L., *Medicago truncatula*-derived calcium oxalate crystals have a negative impact on chewing insect performance via the physical properties. *Entomol. Exp. Appl.*, 2009, **131**(2), 208–215.
 23. Monje, P. V. and Baran, E. J., Characterization of calcium oxalates generated as biominerals in cacti. *Plant Physiol.*, 2002, **128**(2), 707–713.
 24. Hartl, W. P. *et al.*, Diversity of calcium oxalate crystals in Cactaceae. *Can. J. Bot.*, 2007, **85**(5), 501–517.
 25. Nakata, P. A., An assessment of engineered calcium oxalate formation on plant growth and development as a step toward evaluating enhance plant defense. *PLoS ONE*, 2015, **10**(10), e0141982; doi:10.1371/journal.pone.0141982.
 26. Maxwell, D. P. and Bateman, D. F., Influence of carbon source and pH on oxalate accumulation in culture filtrates of *Sclerotium rolfsii*. *Phytopathology*, 1968, **58**, 1351–1355.
 27. Noyes, R. D. and Hancock, J. G., Role of oxalic acid in the *Sclerotinia* wilt of sunflower. *Physiol. Plant Pathol.*, 1981, **18**, 123–132.
 28. Kim, K. S., Min, J. Y. and Dickman, M. B., Oxalic acid is an elicitor of plant programmed cell death during *Sclerotinia sclerotiorum* disease development. *Mol. Plant Microbe Interact.*, 2008, **21**, 605–612.
 29. Dickman, M. B. and de Figueiredo, P., Comparative pathobiology of fungal pathogens of plants and animals. *PLoS Pathog.*, 2011, **7**(12), e1002324.
 30. Kamprath, E. J., Exchangeable aluminum as a criterion for liming leached mineral soils. *Soil Sci. Soc. Am. Proc.*, 1970, **34**, 252–254.
 31. Taylor, G. J., Current views of the aluminum stress response: the physiological basis of tolerance. *Curr. Top. Plant Biochem. Physiol.*, 1991, **10**, 57–93.
 32. Foy, C. D., Soil chemical factors limiting plant growth. *Adv. Soil Sci.*, 1992, **19**, 97–199.
 33. Prasad, R. and Power, J. F., *Soil Fertility Management for Sustainable Agriculture*, CRC-Lewis, Boca Raton, FL, USA, 1997, p. 356.
 34. Morita, A., Yanagisawa, O., Takatsu, S., Maeda, S. and Hiradate, S., Mechanism for the detoxification of aluminum in roots of tea plant (*Camellia sinensis* (L.) Kuntze). *Phytochemistry*, 2008, **69**, 147–153.
 35. Morita, A., Yanagisawa, O., Maeda, S., Takatsu, S. and Ikka, S., Tea plant (*Camellia sinensis* L.) roots secrete oxalic acid and caffeine into medium containing aluminum. *Soil Sci. Plant Nutr.*, 2011, **57**, 796–802.
 36. Ma, J. F., Hiradate, S. and Matsumoto, H., Detoxifying aluminum with buckwheat. *Nature*, 1997, **390**, 569–570.
 37. Ma, J. F., Hiradate, S. and Matsumoto, H., High aluminum resistance in buckwheat. Oxalic acid detoxifies aluminum internally. *Plant Physiol.*, 1998, **117**, 753–759.
 38. Fomina, M., Hillier, S., Charnock, J. M., Melville, K., Alexander, I. J. and Gadd, G. M., Role of oxalic acid over excretion in transformations of toxic metal minerals by *Beauveria caledonica*. *Appl. Environ. Microbiol.*, 2005, **71**, 371–381.
 39. Leitenmaier, B. and Küpper, H., Compartmentation and complexation of metals in hyperaccumulator plants. *Front Plant Sci.*, 2013, **4**, 374; doi:org/10.3389/fpls.2013.00374.
 40. Boyd, R. S., Davis, M. A., Wall, M. A. and Balkwill, K., Nickel defends the South African hyperaccumulator *Senecio coronatus* (Asteraceae) against *Helix aspersa* (Mollusca: Pulmonidae). *Chemoecology*, 2002, **12**, 91–97.
 41. Tao, Q., Hou, D. and Li, T., Oxalate secretion from the root apex of *Sedum alfredii* contributes to hyperaccumulation of cadmium. *Plant Soil*, 2016, **398**(1), 139–152.
 42. McBride, M. B., Richards, B. K., Steenhuis, T., Russo, J. J. and Sauvé, S., Mobility and solubility of toxic metals and nutrients in soil fifteen years after sewage sludge application. *Soil Sci.*, 1997, **162**, 487–500.
 43. Buchauer, M. J., Contamination of soil and vegetation near a zinc smelter by zinc, cadmium, copper, and lead. *Environ. Sci. Technol.*, 1973, **7**, 131–135.
 44. Ghosh, M. and Singh, S. P., A review on phytoremediation of heavy metals and utilization of its by-products. *Appl. Ecol. Environ. Res.*, 2005, **3**(1), 1–18.
 45. Mitikova, T., Prentovic, T. and Markoski, M., Phytoremediation of soils contaminated with heavy metals in the vicinity of smelters for lead and zinc in Velas. *Agric. Conspec. Sci.*, 2015, **80**(1), 53–57.
 46. Wuana, R. A. and Okieimen, F. E., Heavy metals in contaminated soils: a review of sources, chemistry, risks and best available strategies for remediation. *ISRN Ecol.*, 2011, 402647; <http://dx.doi.org/10.5402/2011/402647>.
 47. Tripathi, R. D. *et al.*, Arsenic hazards: strategies for tolerance and remediation by plants. *Trends Biotechnol.*, 2007, **25**(4), 158–165.
 48. Mirza, N., Mahmood, Q., Shah, M. M., Parvez, A. and Sultan, S., Plants as vectors to reduce environmental toxic content. *Sci. World J.*, 2014, 921581, p. 11.
 49. Ng, J. C., Wang, J. and Shraim, A., A global health problem caused by arsenic from natural sources. *Chemosphere*, 2003, **52**(9), 1353–1359.
 50. Chen, C. J., Chen, C. W., Wu, M. M. and Kuo, T. L., Cancer potential in liver, lung, bladder and kidney due to ingested arsenic in drinking water. *Br. J. Cancer*, 1992, **55**(5), 886–892.
 51. Chintakovid, W., Visoothivisthy, P., Khokialtiwong, S. and Lauengsuchonkul, S., Potential of the hybrid marigolds for arsenic phytoremediation and income generation of remediators in Ron Phibun District, Thailand. *Chemosphere*, 2008, **70**(8), 1532–1537.
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